Rearrangement of Zaragozic Acid A Derivatives

Narindar N. Girotra^{*}, Robert A. Reamer, and Mitree M. Ponpipom. Merck Research Laboratories P. O. Box 2000, Rahway, New Jersey, 07065

Abstract: The C6 mesylate of desacyl zaragozic acid A tris tBu ester has been observed to undergo rearrangement in the presence of CsF in DMF.

Zaragozic Acid A (1) is a member of a family of natural products recently reported by two independent groups.^{1,2} These novel compounds are very potent squalene synthase inhibitors.^{1,3} Recently, we described a chemoselective removal and replacement of the C4' and C6 acyl groups of the natural product.⁴ In this paper we report the rearrangement of zaragozic acid A derivatives **3** and **4**.





As part of a program aimed at the preparation of zaragozic acid A analogs, we were interested in replacing the C6 ester group with nitrogen bearing substituents. The mesylate 3 was prepared from the diol 2^4 in 92% yield, and then treated with 15 eq NaN3 in DMF to give new products (one spot by TLC) in *ca* 70% yield. The IR indicates the absence of the azido group. The NMR, beside showing the absence of the mesylate group, reveals the products to be composed of 2 isomeric rearranged compounds (**7a** and **7b** in a ratio *ca* of 2:1). The mass spectrum [C37H52O12; *m*/z 711, (M+Na)+] is consistent with the proposed structure. Further details of the structure determination will be described shortly.



In an attempt to prepare the C6-C7 epoxide, compound 4 prepared from 3 quantitatively, was treated with CsF (10 eq) in DMF at 70 °C for 3 h to give four rearrangend products, which are assumed to arise *via* a Grob-type fragmentation path⁵ initiated by the C4-OH group followed by recyclization in a different mode to give a fused ring system. A detailed examination of this reaction reveals that it affords a mixture of two major kinetic products (polar spot; **6a** : **6b** *ca* 1 : 1) and two minor thermodynamic products (mobile spot, **7a** : **7b** *ca* 2 : 1) in >90% yield. The thermodynamic products were identical with rearrangement products obtained from 3 by treatment with NaN3 in DMF mentioned previously. The ratio of the kinetic and thermodynamic products is *ca* 3 : 1 as estimated by TLC. The kinetic isomers were observed to transform partially to the thermodynamic isomers during preparative TLC, and quantitatively upon treatment with Et3N in ethyl ether or methylene chloride at ambient temperature. However, the kinetic isomers were separated, essentially free from the thermodynamic isomers, by fast flash column chromatography (silica gel; hexanes:ethyl acetate; v/v, 1:4). The isomerization of 6 to 7 is assumed to occur *via* ring opening to give the acyclic intermediate 5 which isomerizes at C3 followed by recyclization. The ratio of the four isomers changed upon standing in NMR solvents favoring 7.



Detailed ¹H-¹H and ¹H-¹³C NMR¹⁰ experiments were used to identify the basic skeletal rearrangement, and NOE difference studies along with ¹H and ¹³C spin-spin coupling constants were employed to establish the stereochemistry and ring conformations. The following discussion is for the major thermodynamic isomer **7a**, but also applies to the other three isomers. One-bond ¹H-¹³C correlation data indicates one of the two oxygen bearing carbon (C6 and C7) in the starting material

now appears to be part of an olefinic system. The proton at 5.90 ppm (H7, doublet, J = 2.6 Hz) was correlated to C7 at 108.5 ppm. The one-bond coupling constant for this carbon was 183.9 Hz, consistent with the dihydrofuran system shown.⁶ Further support for this unsaturation was obtained from the upfield shift (*ca* 4 ppm) of the ester carbonyl at C6. The adjacent carbon (C7a, correlated to H7a at 4.86 ppm) was still at a chemical shift (75.4 ppm) consistent with an oxygen bearing carbon. Long-range heteronuclear correlation data, obtained in an HMBC experiment,⁷ showed all expected two and three bond correlations including the correlation from H7a to C2, consistent with the new ring system.

With the basic skeleton in hand, NOE difference results and coupling constants were used to assign the stereochemistry at C2 and C3. One key piece of data was a four-bond spin-spin coupling between H₃ and the hydroxyl at C₂ (${}^{4}J$ = 1.4 Hz) best explained by a "W" pathway between these protons.⁸ This coupling can only be present if H3 and the hydroxyl are diaxial. NOE difference studies show a 1.4% enhancement from H7 to H3 while the small enhancement from H7a to H3 was ~0.5% thus also supporting the structure as shown. A Dreiding model suggests a small NOE between H_{7a} and H3 or none at all. In 7b the same enhancements from H7 and H7a were observed to H3 supporting the same stereochemistry at C3. Strong support comes from the long-range ¹H-1³C coupling where in 7a the 3J H3-2CO is 1.0 Hz while in 7b the same three-bond path yields a 5.6 Hz coupling, the latter consistent with a diaxial orientation between H3 and the C2-ester carbonyl.⁹ The lack of the four-bond coupling between H3 and the C2 hydroxyl was also consistent with an equatorial orientation of this hydroxyl. Similar arguments can be used to establish the stereochemistry in one of the kinetic isomers (6a) where a four-bond coupling $(^{4}J=1.6 \text{ Hz})$ between H3 and the C2-OH supports their diaxial orientation. An NOE enhancement of 2.2% from H7a to H3 and no observable enhancement from H7 to H3 supports the stereochemistry in 6a as shown. Unlike the thermodynamic isomers, where the six-membered ring in both isomers exists in a chair form, the sixmembered ring in 6b appears to exist in a boat form. In this isomer the NOE enhancement from H7a to H3 is also 2.2%, but a large three-bond ¹H-¹³C coupling from H3 to the C2-carbonyl (as seen in 7b) is not present. The small coupling of 1.6 Hz from H3 to the C2-carbonyl and steric hindrance between C7-C7a and C2-CO2tBu would support a boat conformation in 6b.

In summary, 3, and particularly 4 were found to undergo facile fragmentation followed by ring closure to give four rearranged isomeric products.

Acknowledgments: We thank Mr. R. L. Bugianesi, Dr. G. D. Berger, and Prof. B. M. Trost for helpful discussions. Thanks are also due Ms. M. B. Hill for numbering the compounds 6 and 7 according to the CAS guidelines.

References and Notes:

 Bergstrom, J. D.; Kurtz, M. M.; Rew, D. J.; Amend, A. M.; Karkas, J. D.; Bostedor, R. G.; Germerhausen, J. I; Bansal, V. S.; Dufresne, C.; VanMiddlesworth, F. L.; Hensens, O. D.; Liesch, J. M.; Zink, D. L.; Wilson, K. E.; Onishi, J.; Millilgan, J. A.; Bills, G.; Bartizal, K. F.; Rozdilsky, W.; Abruzzo, G. K.; Kaplan, L.; Nalin, M.; Jenkins, R. G.; Huang, L.; Meinz, M. S.; Quin, L.; Burg, R. W.; Kong, Y. L.; Mochales, S.; Mojena, M.; Martin, I.; Palaez, F.; Diez, M.; Alberts, A. W. Proc. Natl. Acad. Sci. USA, 1993, 90, 80.

- Dawson, M. J.; Farthing, J. E.; Marshall, P. S.; Middleton, R. F. O'Neil. M. J.; Shuttleworth, A.; Stylli, C.; Tait, R. M.; Taylor, P. M.; Wildman, H. G.; Buss, A. D.; Langley, D.; Hayes, M. V. J. Antibiotics 1992, 45, 639.
- Baxter, A.; Fitzgerald, B. J.; Hutson, J. L.; McCarthy, A. D.; Motteram, J. M.; Ross, B. C.; Sepra, M.; Snowden, M. A.; Watson, N. S.; Williams, R. J.; Wright, C. J. Biol. Chem. 1992, 267, 11705.
- 4. Burk, R. M.; Berger, G. D.; Bugianesi, R. L.; Girotra, N. N.; Parsons, W. H.; Ponpipom, M. M. Tet. Lett., 1993, 34, 975.
- 5. Grob, C. A.; Baumann, W. Helv. Chim. Acta, 1955, 38, 594.
- Kalinowski, H.-Q.; Berger, S; Braun, S. in *Carbon-13 NMR Spectroscopy*, John Wiley & Sons, 1988.
- 7. Bax, A.; Summers, M.F. J. Am. Chem. Soc. 1986, 108, 2093.
- 8. Jackman, L. M.; Sternhell, S. in *Applications of NMR Spectroscopy in Organic Chemistry, 2nd. Edition, Pergamon Press*, **1969**.
- 9. Marshall, J. L.; in Carbon Carbon and Carbon Proton NMR Couplings; Applications to Organic Stereochemistry and Conformational Analysis, Verlag Chemie International, 1963.
- 10. NMR data were recorded on a Bruker AM-400 and a Varian XL 400. Diagnostic ¹H and ¹³C assignments are given.

Compound **3** ¹H NMR (CDCl₃): δ 5.95 (d, J=1.5, H₆), 5.10 (d, J=5.0, H₄'), 4.95 (s, H₃), 4.30 (d, J=1.5, H₇), 4.04 (s, 4-OH), 3.25 (s, 3H), 3.05 (s, 3H), 2.05 (s, 3H), 1.42 (s, 3H), 1.34 (s, 3H), , 0.77 (d, J=7.0, 3H).

Compound 4 ¹H NMR (CDCl3): δ 5.85 (d, J=2.0, H₆), 5.10 (d, J=4.5, H₄'), 4.93 (s, H₃) 4.30 (dd, J=4.0, 2.0 H₇), 3.95 (bs, 4-OH), 3.0 (s, 3H), 2.70 (d, J=4.0, 7-OH), 2.0 (s, 3H), 0.79 (d, J=7.0, 3H).

Compound **6a** ¹H NMR (CD₃CN): δ 5.90 (d, J =2.6, H7), 5.12 (d, J =1.4, 2-OH), 4.69 (d, J =2.6, H7_a), 4.58 (d, J =1.4, H₃); ¹³C NMR (CD₃CN): 168.1 (2-CO₂R), 166.0 (3-CO₂R), 159.6 (6-CO₂R), 150.1 (C₆), 112.4 (C₇) 108.4 (C₄₈), 92.8 (C₂), 77.4 (C₇₈), 73.0 (C₃).

Compound **6b** ¹NMR (CD₃CN): 5.85 (d, J = 2.9, H7), 4.94 (s, 2-OH), 4.56 (d, J = 2.9, H7_a), 4.16 (s, H₃); ¹³C NMR (CD₃CN): 167.1₆ (2-CO₂R), 167.1₅ (3-CO₂R), 159.5 (6-CO₂R), 152.4 (C₆), 109.6 (C₇) 109.0 (C_{4a}), 91.6 (C₂), 73.9 (C₃), 73.4 (C_{7a}); characterized as a mixture of **6a** and **6b**.

Compound **7a** ¹H NMR (CD₃CN): δ 5.83 (d, J=2.6, H7), 5.10 (d, J=1.4, 2-OH), 4.86 (d, J=2.6, H7_a), 4.68 (d, J=1.4, H₃); ¹³C NMR (CD₃CN): δ 168.7 (2-CO₂R), 165.7 (3-CO₂R), 159.3 (6-CO₂R), 150.8 (C₆), 108.5 (C₇), 108.3 (C4_a), 92.7 (C₂), 75.4 (C7_a), 71.0 (C₃).

Compound **7b** ¹H NMR (CD₃N): 5.88 (d, J = 2.6, H₇), 5.08 (s, 2-OH), 4.92 (d, J = 2.6, H_{7a}), 4.22 (s, H₃); ¹³C NMR (CD₃CN): 168.3 (2-CO₂R), 166.8 (3-CO₂R), 159.2 (6-CO₂R), 150.7 (C₆), 111.3 (C₇), 108.3 (C_{4a}), 93.6 (C₂), 75.7 (C_{7a}), 72.0 (C₃); characterized as a mixture of **7a** and **7b**.

(Received in USA 10 March 1993; accepted 11 May 1993)